

# The Role of Airway Remodeling in the Pathophysiology and Treatment of Severe Asthma

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Received Date: February 20, 2021; Accepted Date: April 14, 2021; Published Date: May 08, 2021

**Citation:** Nightingale Syabbalo. The Role of Airway Remodeling in the Pathophysiology and Treatment of Severe Asthma. J On Med Bio. 2021;1(1):1002.

## Abstract

Asthma is a highly prevalent chronic inflammatory airway disease, affecting more than 358 million individuals globally, and its prevalence is increasing in many countries. Asthma manifests with respiratory symptoms such as cough, wheezing, dyspnea, and chest tightness that vary over time and in intensity and with variable airflow limitation. Airflow obstruction in patients with asthma is due to airway inflammation, Airway Hyperresponsiveness (AHR), and remodeling which lead to bronchoconstriction in response to allergens, respiratory infections, pollutants, and environmental smoke. Severe asthma is characterized by airway remodeling due to thickening and shedding of the airway epithelium, deposition of Extracellular Matrix (ECM) proteins, subepithelial fibrosis, and Airway Smooth Muscle (ASM) cells hyperplasia and hypertrophy. Immune cells, such as T helper type 2 lymphocytes (Th2), Th17 cells, and innate lymphoid group two cells release multiple cytokines, chemokines, and growth factors which orchestrate airway inflammation, AHR, and activation of structural cells, such as fibroblasts, myofibroblasts, and ASM cells. Structural cells are proliferative and highly secretory. Upon activation, they release a myriad of cytokines, chemokines, adhesion molecules, enzymes, and growth factors, which further promote AHR, and remodeling. Fibroblasts, myofibroblasts, and ASM cells

produce ECM proteins which lead to thickening of the reticular basement membrane, and subepithelial fibrosis cells. Hyperplasia and hypertrophy of the ASM cells result in increase in ASM mass which aggravates airway narrowing and bronchoconstriction. The airway structural changes result in persistent airflow obstruction, and severe asthma. The standard of care treatment, including most of the biologics do not ameliorate airway remodeling. The therapeutic option for severe asthma associated with airway remodeling is to use biologics which target the IL-4/IL-13 and IL-4Ra immune pathway, alarmin cytokines blockers, or bronchial thermoplasty to remove excessive ASM mass.

**Keywords:** Severe asthma; Airway smooth muscle; Airway remodeling; Biologics; Bronchial thermoplasty

## Introduction

Asthma is a highly prevalent chronic inflammatory airway disease, affecting more than 358 million individuals globally, and its prevalence is rising in many countries. The Global Initiative for Asthma (GINA) defines asthma as a heterogenous disease usually characterized by chronic airway inflammation [1]. It manifests with respiratory symptoms such as cough, wheezing, dyspnoea, and chest tightness that vary over time and in intensity and with variable airflow limitation [1]. The variable airflow limitation in patients with asthma is mainly due to

contraction of the hypertrophic and hypercontractile Airway Smooth Muscle (ASM) cells leading to bronchoconstriction in response to allergens, respiratory infections, pollutants, environmental tobacco smoke, and chemical irritants.

Airway obstruction in asthmatics is also due to thickening of the reticular basement membrane and subepithelial fibrosis; deposition of Extracellular Matrix (ECM) proteins; goblet cells hyperplasia and mucus hypersecretion, angiogenesis, vascular congestion, and mucosal oedema. The complex structural change which occurs in severe asthmatic airways is termed airway remodeling [2,3]. Table 1 lists the pathophysiological mechanisms of airway obstruction in patients with severe asthma.

**Table 1:** Mechanisms of airway remodeling in patients with severe asthma

Epithelial injury due to allergen proteases, pollutants, and respiratory viral infections
Release of cytokines, chemokines, growth factors, and adhesion molecules
Airway epithelial thickening, and shedding
Epithelial cell apoptosis, and release of “alarmin” cytokines
Submucous glands and goblet cell hyperplasia, and mucus hypersecretion
Activation of myofibroblasts, and fibroblasts
Deposition of extracellular matrix proteins
Reticular basement membrane thickening and subepithelial fibrosis
Mast cell infiltration of airway smooth muscle cells
Airway smooth muscle hyperplasia, and hypertrophy
Neoangiogenesis, and exaggerated vasodilatation
Airway hyper responsiveness
Airway remodeling
Corticosteroid-resistance

Airway smooth muscle cells play a key role in modulating airway caliber in patients with asthma. Airway remodeling in asthmatic airways is characterized by hyperplasia and hypertrophy of ASM cells which contribute to severe airway obstruction. Additionally, ASM cells from patient with asthma are dysfunctional. They display plasticity, and take on the hypercontractile, proliferative, and secretory

phenotype upon priming by inflammatory mediators [4-6]. Activated ASM cells release proinflammatory cytokines, chemokines, adhesion molecules, and lay down Extracellular Matrix (ECM) proteins, which amplify the inflammatory responses, and perpetuate airway hyperresponsiveness, and airway remodeling [7-10]. The ASM cells produce chemokines, such as CXCL10 which is a potent chemoattractant for mast cells, and T lymphocytes. CRCL10 via its receptor CXCR3 and other chemokines promote migration of mast cells into ASM bundles, here after, activated mast cells release a plethora of granule preformed and newly synthesized mediators, cytokines, chemokines, growth factors, and enzymes [10-12]. The mast cell-derived inflammatory mediators further orchestrate airway inflammation, AHR, and airway remodeling, resulting in severe asthma.

Goblet cells hyperplasia, and mucus hypersecretion is a prominent feature of asthma. Patients with asthma secrete pathological mucus composed of mostly MUC5AC mucin which is tenacious, and sticky to the mucus secreting cells. Excessive mucilaginous mucus leads to airways mucus plugging, and is associated with severe asthma, and fatal asthma [13].

Angiogenesis and expansion of the bronchial vasculature with vasodilated permeable blood vessels accompanies airway remodeling in asthma. It is associated with mucosal edema, and airway inflammation due to leakage of cytokines and chemokines; and diapedesis of inflammatory cells from the dilated blood vessels.

Treatment of severe asthma due to ASM dysfunction and airway remodeling is challenging. The Standard of Care (SoC) treatment, including high-dose Inhaled Corticosteroids (ICS), and most biologics do not ameliorate airway remodeling, especially due to ASM cells hyperplasia and hypertrophy, and subepithelial fibrosis. The therapeutic option for severe asthma associated with severe remodeling is probably to administer biologics which target the IL-4/IL-13 and IL-4R $\alpha$  immunological pathway, and “alarmin” monoclonal Antibodies (mAb), or bronchial thermoplasty to remove excessive ASM mass [14].

## Airway Remodeling in Severe Asthma

Severe asthma is characterized by active airway inflammation, Airway Hyperresponsiveness (AHR), and remodeling. Airway remodeling is a complex pathophysiological process involving structural changes, such as ASM cells hypertrophy and migration, reticular basement membrane thickening and subepithelial fibrosis submucous gland and goblet cells hyperplasia thickening and shedding of the airway epithelium, and epithelial cells apoptosis [15-30]. Airway remodeling is also accompanied by neovascularization, and expansion of the airway vascular bed, vasodilatation, and mucosal oedema [31,32]. All of the above structural changes lead to airway narrowing, excessive bronchoconstriction, and severe, uncontrolled asthma. Table 1 summarizes the pathophysiologic mechanisms of airway remodeling in patients with severe asthma.

## Extracellular Matrix Proteins

Airway remodeling involves laying of Extracellular Matrix (ECM) proteins by structural cells, principally fibroblasts, and to a lesser extent myofibroblasts, and ASM cells. The ECM proteins are composed of several proteins and glycoproteins, such as collagens I, II, III, V, and XI; adhesion molecules including fibronectin and tenascin-C; proteoglycans (lumican and biglycan); and glycosaminoglycans [33-37]. The ECM protein is produced mostly by fibroblasts, and to lesser extent by myofibroblasts, and ASM cells [35,37,38-41]. The ECM proteins provide structural and mechanical support of the airways, and provide a conducive milieu for cell adhesion, migration, proliferation, and activation [24]. Nevertheless, the structural changes lead to narrow and stiff airways, and correlate with clinical and functional severity of asthma [21,42,43]. Airway remodeling is seen in early stages of asthma, but correlates with the duration of severe asthma, and is associated with corticosteroid resistance [44-46].

## Airway Mucus Secretion in Asthma

Airway remodeling in asthma is accompanied by goblet cells, and submucous glands hypertrophy and hyperplasia, mucus hypersecretion, and abnormal mucus [47]. The epithelial dysfunction in asthma [48,49], leads to impaired

mucociliary transport, accumulation of mucus, and mucus plugging of the airways [47,50,51]. Dysfunctional mucociliary escalator is even evident in mild asthma, and decreases further during acute exacerbations [52-54]. Failure to clear mucus expectoration has been associated with death due to asphyxiation from intraluminal airway obstruction due to mucus plugs [55-57].

Humans secrete five types of gel-forming mucins (MUC2, MUC6, MUC19, MUC5AC, and MUC5B [25], but the mucins secreted in large quantities, and responsible for the viscoelastic properties of mucus depend on MUC5AC, and MUC5B [50]. Genome World-Wide Studies (GWAS) have revealed that polymorphism of 11p15 MUC5B, and MUC5AC locus is associated with airway hyper responsiveness, and asthma [58]. Altered expression of the pathologic mucin MUC5AC potentially contributes to mucus plugging and airway obstruction [59]. Furthermore, many patients with asthma have increased MUC5AC mRNA levels, but decreased MUC5B levels, thus the composition of mucus contributes to the viscoelasticity, and mucilaginous nature of mucus [60].

One of the characteristics mucus secreted by asthmatic patients is the change in the relative proportion of MUC5AC and MUC5B, and the organization of airway mucus which is likely to affect its properties, and contribute to airways mucus plugging [47]. Asthmatic airways produce more MUC5AC than MUC5B. It appears that MUC5AC is the mucin which is responsible for the mucilaginous mucus which is difficult to clear, because of the tethering of MUC5AC-containing mucus domains to mucus-producing cells in the epithelium [61]. Accumulation of pertinacious mucus causes diffuse airway obstruction, and is an important feature of severe, near fatal, and fatal asthma [55,56,60].

Interleukin-13 (IL-13 is a Th2 cytokine, produced by Type 2 helper (Th2) cells, and Innate Lymphoid Group 2 Cells (ILC2). It plays a key role in the production and secretion of excessive mucus from goblet cells, and submucous glands in patients with asthma. IL-13 has been shown to induce significant increases in the expression of MUC5AC in human epithelial cells in vitro, and in murine asthma model

[62,63-66]. Additionally, IL-13 dramatically impairs mucociliary transport of mucus containing MUC5AS mucus domain, and is responsible for other immunopathological features of severe eosinophilic asthma, such as subepithelial fibrosis [67]. Theoretically, targeting IL-13 and its sub-receptor IL-4R $\alpha$  is a potential therapeutic option to ameliorate mucus hypersecretion and plugging.

Epidermal Growth Factor Receptor (EGFR) signaling is required for mucus production in vitro and vivo [65,68-70]. It induces MUC5AC expression [69], and EGFR levels are significantly increased in patients with asthma, correlating with the severity of the disease [70].

Pathological mucus and dysfunctional mucus escalator can lead to mucus plugging, and airflow obstruction in severely bronchoconstrictor airways. Currently, there are no effective mucolytic or mucoregulator drugs for the treatment of airway obstruction due to sticky mucus in patients with asthma [71,72]. The role of biologics targeted against Th2 cytokines has not yet been established as mucolytic or mucoregulator agents. MUC5AC targeted therapies might offer the opportunity to ameliorate mucus plugging and airway obstruction in patients with asthma [47].

### Airway Smooth Muscle Hyperplasia and Hypertrophy

Airway smooth muscle cell hyperplasia and hypertrophy is a cardinal feature of airway remodeling in asthma. It is a major determinant of bronchoconstriction in response to allergen, respiratory viral infections, pollutants, and environmental tobacco smoke. Increase in ASM mass occurs early in the development of asthma [73], and is present even in school children with asthma [74-76], and in patients with mild-to-moderate asthma [77]. The increase in ASM mass occurs in most of the phenotypes of asthma, including neutrophilic asthma, and paucigranulocytic asthma [14]. The degree of increase in ASM hyperplasia, and ASM cell structural and phenotypic change correlate with the severity of asthma [78-82], and the duration of severe asthma in older patients [83]. Furthermore, severe ASM cell hyperplasia and hypertrophy has been associated with status asthmaticus, and fatal asthma [25,83-87].

Airway smooth muscle cell hypertrophy and hyperplasia is mostly due to the profibrotic and proliferative effects of growth factors, such as TGF- $\beta$  [88-90], and due to infiltration of the muscle bundles by myofibroblasts, and neighboring ASM cells migration from the submucosa compartment, or hematopoietic progenitor cells from the circulation [91-94]. TGF- $\beta$  plays an important role in the increase in ASM mass and hypercontractility. It is responsible for activation of the Reactive Oxygen Species (ROS)-generating enzyme (NADPH oxidase 4 (Nox 4) that induces myofibroblasts differentiation [95], and is implicated in ASM cell proliferation, and hyper contractility [96,97]. The mechanisms by which myofibroblast transform to ASM cell is speculative but probably represent part of the spectrum of plasticity of mesenchymal cells through transition from fibroblast to myofibroblast, and finally into ASMS cells [98]. Wicks, et al. [99] have reported that TGF- $\beta$ 2 may enhance upregulation of ASM cell from asthmatic myofibroblasts.

Another possible source of increase in the ASM cell numbers would be from differentiation of tissue resident Mesenchymal Stem Cells (MSCs); or from Epithelial-Mesenchymal Transition (EMT) of epithelial cells [100]. Additionally, pericytes transformation and migration may also contribute to the increase in ASM mass [98]. However, in order to transfer to functional ASM cells or myofibroblasts, all these cells require loss of their non-mesenchymal markers and acquire mesenchymal characteristics, such increase in mesenchymal proteins, including  $\alpha$ -smooth muscle actin ( $\alpha$ -ASMA), N-cadherin, and vimentin expression [98]. Not with standing, fibroblasts, myofibroblasts, and ASM cells from asthmatic patients display tremendous plasticity.

Migration of ASM cells from the submucosa and from other ASM bundles, may partly explain the increase in ASM mass in patients with asthma. Multiple lipid mediators, such as leukotrienes (LTE4, and LTB4), and prostaglandins (PGD2) [92] have been shown to stimulate ASM cell migration into muscle bundles. Similarly multiple cytokines, such as IL-8, IL-13, IL-17, IL-25, IL-33, TSLP, and TNF- $\alpha$  [101-104], chemokines (CXCL2, CXCL3, CXCL8, CCL19, eatoxins,

and RANTES) [101,105-107], and growth factor, including platelet-derived growth factor, TGF- $\beta$  [92,93] stimulate ASM cells migration. Chemokines, cytokines, and growth factors synergize with other chemo attractants, such as CXCL8 (IL-8), eotaxins, and RANTES in order to promote ASM cells migration. Notably, activated ASM cells in vitro are capable of releasing cytokines, such as IL-5, IL-6, and IL-8, and chemokines, including CCL5, CXCL8, and eotaxins [108-109], which may act in a positive feedback autocrine fashion to enhance ASM cells migration and contraction [106,108,110]. Inflammatory mediators which promote ASM cells proliferation and migration are shown in Table 2.

**Table 2:** Airway smooth muscle cells proliferative and migratory mediators in asthma.

Cytokines	Chemokines
Interleukin (IL) IL-1 $\beta$	CCL2 (MCP-1, 3, 5)
IL-2	CCL5 (RANTES)
IL-5	CCL11 (eotaxin)
CCL10	CXCL10
IL-6	CX <sub>3</sub> CL1 (Fraktalkine)
IL-8	CXCL8 (IL-8)
IL-10	Growth factors
IL-11	TGF- $\beta$ 1
IL-12	bFGF-1
TSPL	PDGF-BB
Lipid mediators	
Leukotriene D4 (LTD4)	VEGF
Thromboxane A2	SCF
Enzymes	TNF- $\alpha$
Tryptase	CTGF
$\beta$ -Hexosaminidase	IGF-1, IGF-2
Lysosomal hydroxylase	
MMP-9, MMP-12	Neuropeptide transmitters
Thrombin	Bradykinin
Small-molecule transmitters	Angiotensin II
Histamine	Endothelin-1
Serotonin	

Angiogenesis is a prominent feature of airway remodeling in patients with asthma. It is a complex process of formation of new blood vessels, and enlargement and expansion of the existing vessels, induced by angiogenic factors and counteracted by angiostatin factors [51,111]. The subepithelial submucosa of the airways in asthmatic patients is characterized by an increase in the vascular density [112-115]. The total number of blood vessels, and vascular area is increased about two- to three-folds compared to healthy control subjects [116,117]. The blood vessels are dilated and more permeable leading to airway mucosal oedema [118], and inflammation due to leakage of pro-inflammatory mediators, and diapedesis of inflammatory cells through the capillary pores [119]. The increase in airway vasculature has been shown to correlate with airflow obstruction [112,120,121], and airway hyper responsiveness [122].

Angiogenesis is a complex tightly regulated process mediated by a balance between multiple proangiogenic factors and antiangiogenic cytokine, chemokines, growth factors, and enzymes [31,32,123-126]. The proangiogenic factors include growth factors, such as Vascular Endothelial Growth Factor (VEGF), Basic Fibroblast Growth Factor (bFGF), Epidermal Growth Factor (EGF), Insulin-Like Growth Factor (IGF-1), Amphiregulin, Angiopoietin-1, and 2 (Ang-1, Ang-2), and angiogenin [124]. The cytokines involved in angiogenesis include IL-6, IL-8 [122], and IL-25 [126]. The most potent profibrotic growth factor is VEGF, which has six splice-variants [127]. VEGF promotes enlargement of existing vascular structures from existing ones, by causing endothelial cell proliferation and migration [127,128]. It also stimulates formation of new capillaries, and blood vessels [127,128]. Additionally, VEGF significantly increases the permeability of blood vessel, almost about 50,000 times more potent than histamine [129]. VEGF concentration in sputum and in lung tissue is elevated, and correlates with the severity of asthma [130]. Co-expression of VEGF with Ang-1, and Ang-2 has been shown to induce proliferation and migration of endothelial cells, and sprouting of new capillaries, and enlargement of existing capillaries [130]. Ang-1 stimulates migration of pericytes and ASM cells and therefore stabilizes the new

## Angiogenesis in Asthma

capillary tubes during angiogenesis [131]. Angiogenesis is further enhanced by secretion of proangiogenic factors by activated ASM cells, such as VEGF, angiogenin, and angiopoietin-1 [111,132], thus perpetuating neovascularization.

Airway angiogenesis is counterchecked by multiple antiangiogenic factors, such as arrestin tumstatin, canstatin, endostatin, and arrestin [124]. However, in patients with asthma, the proangiogenesis mechanisms override antiangiogenic factors in the vascular remodeling process. Other asthma-associated stimuli, such as polymorphism in the ADAM33 gene [133], environmental tobacco smoke [134], and rhinovirus infection also promote angiogenesis [134].

Neovascularization and vasodilatation lead to increased vascular permeability, and mucosal oedema which narrows the airway lumen. Furthermore, transmigration of inflammatory cells, and leakage of mediators result in more airway inflammation, AHR, and airway remodeling. Increase in the submucosal microvascular plexus, vasodilatation, and mucosal oedema correlate with asthma severity [112,116,123], and has been associated with fatal asthma [111, 135].

### **Treatment of Airway Remodeling in Severe Asthma**

Treatment of airway remodeling in patients with severe asthma due to airway remodeling is challenging. Currently, there is no specific pharmacological treatment for airway remodeling, including biologics, with the potential exception of IL-4/IL-13 targeted therapies, such as dupilumab [136], and probably anti-TSLP, namely tezepelumab [137]. Dupilumab (Dupixent®) is a fully humanized IgG4 monoclonal antibody to the IL-4R $\alpha$ , which mediates signaling to both IL-4 and IL-13, and blocks their immunological pathways and suppressing airway eosinophilic inflammation [138]. Dupilumab is the only biologic which has been approved for the treatment of eosinophilic asthma, allergic rhinitis, chronic rhinosinusitis and nasal polypsis, and atopic dermatitis [139,140], and eosinophilic oesophagitis [141], and is highly effective in treating these conditions. Patients with the above co-

morbidities should be considered for the so called “magic bullet” biologic.

Tezepelumab is a first-in-class fully human IgG2 monoclonal Antibody (mAb) that binds to TSLP, and prevents it to interact with its receptor TSLPR, thus inhibiting multiple downstream immunopathologic pathways, and production of cytokines, and chemokines [142]. Several clinical trials have shown that tezepelumab can reduce both the early and late asthmatic responses due to allergen challenges, and significantly reduce the AAER, and improve lung function and HLQoL in patients with eosinophilic asthma and non-eosinophilic asthma [143-146]. Furthermore, tezepelumab had been demonstrated to reduce eosinophil counts, and eosinophilic biomarkers of airway inflammation, such as serum IgE, and FeNO [144-146]. Due its pathophysiological roles in the pathophysiology of eosinophilic, and neutrophilic asthma, and airway remodeling, tezepelumab might prevent or ameliorate airway remodeling [147]. It is effective as add-on treatment of different phenotypes of asthma irrespective of the baseline biomarkers of airway inflammation.

Brochial Thermoplasty (BT) is a bronchoscopic therapeutic intervention which uses a special AlairTM catheter (Bronchial Thermoplasty System, Natick, MA, USA) to remove excessive ASM mass, ECM proteins, subepithelial fibrosis, sub mucus glands, and nerve endings [148,149]. The criteria for selection of patients for BT, and the procedure are discussed in great detail elsewhere [150-154]. Bronchial thermoplasty has been shown to be relatively safe, and result in significant improvement in asthma control, and HRQoL, and reduction in annual exacerbation rates compared with sham procedure [155]. BT was approved by the U.S. Food and Drug Administration (FDA) in 2010 [156], and is approved in several European Community countries, UK, Australia, Brazil, Canada, Japan, and South Korea for the treatment of severe persistent asthma in patients 18 years and older, that is not controlled by high-dose ICS, and LABA.

### **Conclusion**

Airway remodeling is a complex structural change in the airways due to deposition of ECM proteins, thickening of

the reticular basement membrane, subepithelial fibrosis, goblet cell hyperplasia and mucus hypersecretion, and ASM hyperplasia and hypertrophy. Currently, there are no specific pharmacological agents, and biologics for the treatment of the increase in ASM mass, and airway remodeling in patients with severe asthma. Biologics targeting IL-4/IL-13 and IL-4R $\alpha$  immunological pathway, such as dupilumab, and anti-TSLP, including tezepelumab might ameliorate airway remodeling. Bronchial thermoplasty which removes excessive ASM mass, matrix proteins, and subepithelial fibrosis is a treatment option for highly selected patients in experienced centers.

## References

1. Global Initiative for Asthma. Global Strategy for Asthma management and Prevention. April 2020.
2. James AL, Pare PD, Hogg JC. The mechanics of airway narrowing in asthma. *Am Rev Respir Dis*. 1989; 139(1):242-6.
3. Lemanske RF Jr, Busse WW. Asthma. *J Allergy Clin Immunol*. 2003;111(2Suppl):S502-19.
4. Johnson PR, Burgess JK. Airway smooth muscle and fibroblasts in the pathogenesis of asthma. *Curr Allergy Asthma Rep*. 2004;4(2):102-8.
5. Woodruff PG. Gene expression in asthmatic airway smooth muscle. *Proc Am Thorac Soc*. 2008;5(1):113-8.
6. Bergeron C, Tulic MK, Hamid Q. Airway remodelling in asthma: From benchside to clinical practice. *Can Respir J*. 2010;17(4):e85-e93.
7. Panettieri RA Jr. Airway smooth muscle: An immunomodulatory cell. *J Allergy Clin Immunol*. 2002; 110(Suppl 6):S269-274.
8. Hakanarson H, Maskeri N, Carter C, Grunstein MM. Regulation of TH1- and TH2-type cytokine expression and action in atopic asthmatic sensitized airway smooth muscle. *J Clin Invest*. 1999;103(7):1077-87.
9. Johnson PR. Role of human airway smooth muscle in altered extracellular matrix production in asthma. *Clin Exp Pharmacol Physiol*. 2001;28(3):233-236.
10. Reischl IG, Coward WR, Church MK. Molecular consequences of immunoglobulin E receptor (IgE-FcR1) interaction. *Biochem Pharmacol*. 1999;58(12):1841-50.
11. Page S, Ammit AJ, Black JL, Armour CL. Human mast cell and airway smooth muscle interactions: implications for asthma. *Am J Physiol Lung Cell Mol Biol*. 2001;281(16):L1313-23.
12. Brightling CE, Ammit AJ, Kaur D, Black JL, Wardlaw AJ, Hughes JM, et al. The CXCL10/CXCR3 axis mediates human mast cell migration to asthmatic airway smooth muscle. *Am J Respir Crit Care Med*. 2005;171(10):1103-8.
13. Aikawa T, Shimura S, Sasaki H, Ebina M, Takishima T. Goblet cell hyperplasia with mucus accumulation in the airway of patients who died of severe asthma. *Chest*. 1992;101(4):916-21.
14. Syabbalo N. Clinical features and management of paucigranulocytic asthma. *Ann Clin Med Res*. 2020; 1(3):1011.
15. Huber HL, Koessler KK. The pathology of bronchial asthma. *Arch Intern Med*. 1922;30(6):689-760.
16. Niimi A, Matsumoto H, Amitani R, Nakano Y, Mishima M, Minakuchi M, et al. Airway wall thickness in asthma assessed by computed tomography. Relation to clinical indices. *Am J Respir Crit Care Med*. 2000;162(4 Pt 1):1518-23.
17. Little SA, Sproule MW, Cowan MD, Macleod KJ, Robertson M, Love JG, et al. High resolution computed tomographic assessment of airway wall thickness in chronic asthma: reproducibility and relationship with lung function and severity. *Thorax*. 2002;57(3):247-53.
18. James AL, Bai TR, Mauad T, Abrahamson MJ, Dolhnikoff M, McKay PS, et al. Airway smooth muscle thickness in asthma is related to severity but not duration of asthma. *Eur Respir J*. 2009;34(5):1045.
19. Bara I, Ozier A, J-M Tunon de Lara R, Marthan R, Berger P. Pathophysiology of bronchial smooth

- muscle remodeling in asthma. *Eur Respir J.* 2010;36(5):1174-84.
20. Ozier A, Allard B, Bara I, Girodet P-O, Trian T, Marthan R, et al. The pivotal role of airway smooth muscle in asthma pathophysiology. *J Allergy (Cairo).* 2011;2011:742710.
21. James AL, Elliot JG, Jones RL, Carroll ML, Mauad T, Bai TR, et al. Airway smooth muscle hypertrophy and hyperplasia in asthma. *Am J Respir Crit Care Med.* 2012;185(10):1058-64.
22. Roche WR, Beasley R, Williams JH, Holgate SJ. Subepithelial fibrosis in the bronchi of asthmatics. *Lancet.* 1989;1(8637):520-4.
23. James AL, Maxwell PS, Pearce-Pinto G, Elliot JG, Carroll NG. The relationship of reticular basement membrane to airway wall remodeling in asthma. *Am J Respir Crit Care Med.* 2002;16(12 Pt 1):1590-5.
24. Hough KP, Curtiss ML, Blain TJ, Liu R-M, Trevor J, Deshane JS, et al. Airway remodeling in asthma. *Front Med.* 2020;7:191.
25. Carroll N, Elliot J, James A. The structure of large and small airways in nonfatal and fatal asthma. *Am Rev Respir Dis.* 1993;147(2):405-10.
26. Jenkins HA, Cool C, Szeffler SJ, Covar R, Brugman S, Gelfand EW, et al. Histopathology of severe childhood asthma: a case series. *Chest.* 2003;124(1):32-41.
27. Fahy JV. Goblet cell and Puccini gene abnormalities in asthma. *Chest.* 2002;122(6 Suppl):320S-326S.
28. Holgate ST. The sentinel role of the epithelium in asthma pathogenesis. *Immunol Rev.* 2011;242(1):205-19.
29. Proud D, Leigh R. Epithelial cells and airway diseases. *Immunol Rev.* 2011;242(1):186-204.
30. Loxham M, Davies DE, Blume C. Epithelial function and dysfunction in asthma. *Clin Exp Allergy.* 2014;44(11):299-313.
31. Bischof R, Bourke JE, Hirst SJ. Measurement and impact of remodeling in the lung airway neovascularization in asthma. *Proc Am Thorac Soc.* 2009;6(8):673-7.
32. Ribatti D, Puxeddu I, Crivellato E, Nico B, Vacca A, Levi-Schaffer F. Angiogenesis in asthma. *Clin Exp Allergy.* 2009;39(12):1815-21.
33. Araujo BB, Dolhnikoff M, Silva LF, Elliot J, Lindeman JH, Ferreira DS, et al. Extracellular matrix components and regulators in the airway smooth muscle in asthma. *Eur Respir J.* 2008;32(1):61-9.
34. Royce SG, Chang V, Samuel CS, Tang ML. The regulation of fibrosis in airway remodeling in asthma. *Mol Cell Endocrinol.* 2012;351(2):167-75.
35. Mostaco-Guidolin LB, Osei ET, Ullah J, Hajimohammadi S, Fouadi M, Li X, et al. Defective Fibrillar collagen organization by fibroblasts contributes to airway remodeling in asthma. *Am J Respir Crit Care Med.* 2019;200:431-443.
36. Ito JT, Lourenco JD, Righetti RF, Tiberio I, Prado CM, Lopes F. Extracellular component remodeling in respiratory disease: what has been found in clinical and experimental studies? *Cells.* 2019;8(4):342.
37. Reeves SR, Kolstad T, Lien TY, Elliot M, Ziegler SF, Wight TN, et al. Asthmatic airway epithelial cells differentially regulate fibroblast expression of extracellular matrix components. *J Allergy Clin Immunol.* 2014;134(3):663-670.e1.
38. Brewster CE, Howarth PH, Djukanovic R, Wilson J, Holgate ST, Roche WR. Myofibroblasts and subepithelial fibrosis in bronchial asthma. *Am J Respir Cell Mol Biol.* 1990;3(5):507-11.
39. Cheng W, Yan K, Xie LY, Chen F, Yu HC, Huang YX, et al. MiR-143-3p controls TGF-1-induced cell proliferation and extracellular matrix production in airway smooth muscle via negative regulation of nuclear factor of activated T cells. *Mol Immunol.* 2016;78:133-9.
40. Harkness LM, Weckmann M, Kopp M, Becker T, Ashton AW, Burgess JK. Tumstatin regulates the

- angiogenic and inflammatory potential of airway smooth muscle extracellular matrix. *J Cell Mol Med.* 2017;21(12):3288-97.
41. Koopmans T, Crutzen S, Menzen MH, Halayko AJ, Hackett TL, Knight DA, et al. Selective targeting CREB-binding protein/-catenin inhibits growth of and extracellular matrix remodelling by airway smooth muscle. *Br J Pharmacol.* 2016;173(23):3327-41.
42. Chetta A, Foresi A, Del Donno M, Bertorelli G, Pessi A, Olwieri D. Airway remodeling is distinctive feature of asthma and is related to severity of the disease. *Chest.* 1997;111(4):852-7.
43. Hoshimi M, Nakamura Y, Sim JJ. Expression of growth factors and remodelling of airway wall in bronchial asthma. *Thorax.* 1998;53(1):21-7.
44. James A. Airway remodeling in asthma: is it fixed or variable. *Am J Respir Crit Care Med.* 2017;195(8):968-70.
45. Bousquet J, Jeffery PK, Busse WW, Johnson M, Vignola AM. Asthma. From bronchoconstriction to airway inflammation and remodeling. *Am J Respir Crit Care Med.* 2000;161(5):1720-45.
46. Jeffery PK. Remodeling in asthma and chronic obstructive lung disease. *Am J Respir Crit Care Med.* 2001;164(10 Pt 2):S28-38.
47. Bonser LR, Erle DJ. Airway mucus and asthma: The role of MUC5AC and MUC5B. *J Clin Med.* 2017;6(12):112.
48. Heijink IH, Nawijn MC, Hackett T-L. Airway epithelial barrier function regulates the pathogenesis of allergic asthma. *Clin Exp Allergy.* 2014;44(5):620-630.
49. Loxham M, Davies DE, Blume C. Epithelial function and dysfunction in asthma. *Clin Exp Allergy.* 2014;44(11):299-313.
50. Fanta C. Asthma. *N Engl J Med.* 2009;360(10):1002-14.
51. Dunnill MS. The pathology of asthma with special references to changes in the bronchial mucosa. *J Clin Pathol.* 1960;13(1):27-33.
52. Bateman JR, Pavia, Sheahan NF, Agnew JE, Clarke SW. Impaired tracheobronchial clearance in patients with mild asthma. *Thorax.* 1983;38(6):463-7.
53. O'Riordan TG, Zwang J, Smaldone GC. Mucociliary clearance in adult asthma. *Am Rev Respir Dis.* 1992;146(3):598-603.
54. Messina MS, O'Riordan TG, Smaldone GC. Changes in mucociliary clearance during acute exacerbations of asthma. *Am Rev Respir Dis.* 1991;143(5 Pt 1):993-7.
55. Messer JW, Peter GA, Bennett WA. Causes of death and pathological finding in 304 cases of bronchial asthma. *Dis Chest.* 1960;38:616-624.
56. Kuyper LM, Pare PD, Hogg JC, Lambert R, Ionescu D, Woods R, Bai TR. Characterization of airway plugging in fatal asthma. *Am J Med.* 2003;115(1):6-11.
57. Evans CM, Kim K, Tuvim MJ, Dickey BF. Mucus hypersecretion in asthma: Causes and effects. *Curr Opin Pulm Med.* 2009;15(1):4-11.
58. Jeffery PK, Li D. Airway mucosa: Secretory cells, mucus and Puccini genes. *Eur Respir J.* 1997; 10(7):1655-62.
59. Kabesch M, Jost J. Recent findings in the genetic and epigenetic of asthma and allergy. *Sem Immunopathol.* 2020;42(1):43-60.
60. Aikawa T, Shimura S, Sasaki H, Ebina M, Takishima T. Marked goblet cell hyperplasia with mucus accumulation in the airways of patients who died of severe asthma attack. *Chest.* 1992;101(4):916-921.
61. Bonser LR, Zlock L, Finkbeiner W, Erle DJ. Epithelial tethering of MUC5AC-rich mucus impairs mucociliary transport in asthma. *J Clin Invest.* 2016;126(6):2367-71.
62. Welsh KG, Rousseau K, Fisher G, Bonser LR, Bradding P, Brightling CE, et al. MUC5AC and glycosylated variant of MUC5B alter mucus composition in children with acute asthma. *Chest.* 2017;152(4):771-9.

63. Kuperman DA, Lewis CC, Woodruff PG, Rodriguez MW, Yang YH, Dolganov GM, et al. Dissecting asthma using focused transgenic modeling and functional genomics. *J Allergy Clin Immunol.* 2005;116(2):305-11.
64. Zhen G, Park SW, Nguyenvnu LT, Rodriguez MW, Barbeau R, Paquet AC, et al. IL-13 and epidermal growth factor have critical but distinct roles in epithelial cell Puccini production. *Am J Respir Cell Mol Biol.* 2007;36(2):244-53.
65. Kuperman DA, Huang X, Koth LL, Chang GH, Dolganov GM, Zhu Z, et al. Direct effects of interleukin-13 on epithelial cells cause airway hyperreactivity and mucus overproduction in asthma. *Nat Med.* 2002;8(8):885-9.
66. Laoukili J, Perrett E, Willems T, Minty A, Parthoens E, Houcine O, et al. IL-13 alters mucociliary differentiation and ciliary beating of human respiratory epithelial cells. *J Clin Invest.* 2001;108(12):1817-24.
67. Fichtner-Feigl S, Strober W, Kawakami K, Puri RK, Kitani A. IL-13 signaling through IL-13 alpha 2 receptor is involved in induction of TGF-beta 1 production and fibrosis. *Nat Med.* 2006;12(1):99-106.
68. Takeyama K, Dabbagh K, Lee H-M, Augusti C, Lausier JA, Ueki IF, et al. Epidermal growth factor system regulates Puccini production in airways. *Proc Natl Acad Sci USA.* 1999;96(6):3081-6.
69. Cras TDL, Acciani TH, Mushaben EM, Kramer EL, Pastura PA, Hardie WD, et al. Epithelial EGF receptor signaling mediates airway hyperreactivity and remodeling in mouse model of chronic asthma. *Am J Physiol Lung Cell Mol Physiol.* 2011; 300(3):L414-21.
70. Takeyama K, Fahy JV, Nadel JA. Relationship of epidermal growth factor receptors to goblet cell production in human bronchi. *Am J Respir Crit Care Med.* 2001;163(2):511-6.
71. Rogers DF, Barnes PJ. Treatment of airway mucus hypersecretion. *Ann Med.* 2006;38(2):116-25.
72. Sadowska AM. N-Acetylcysteine mucolysis in the management of chronic obstructive pulmonary disease. *Ther Adv Respir Dis.* 2012;6(3):127-135.
73. Bousquet J, Jeffery PK, Busse WW, Johnson M, Vignola AM. Asthma. From bronchoconstriction to airway inflammation and remodeling. *Am J Respir Crit Care Med.* 2000;161(5):1720-45.
74. Regamey N, Ochs M, Hillard TN, Muhrfeld C, Cornish N, Fleming L, et al. Increased airway smooth muscle in children with asthma, cystic fibrosis and bronchiectasis. *Am J Respir Crit Care Med.* 2008;177(8):837-43.
75. Tillie-Leblond I, de Blic J, Jaubert F, Wallaert B, Scheimann P, Gosset P. Airway remodeling is correlated with obstruction in children with severe asthma. *Allergy.* 2008;63(5):533-41.
76. O'Reilly R, Illman N, Irving S, Bossley CJ, Sonnappa S, Zhu J, et al. Increased airway smooth muscle in preschool wheezers who have asthma at school age. *J Allergy Clin Immunol.* 2012;131(4):1024-32.
77. Woodruff PG, Dolganor GM, Ferrado RE, Donnelly S, Hays SR, Soberg OD, et al. Hyperplasia of smooth muscle in mild to moderate asthma without changes in cell size or gene expression. *Am J Respir Crit Care Med.* 2004;169(9):1001-6.
78. Pepe C, Foley S, Shannon J, Lemiere C, Olivenstein R, Ernst P, et al. Differences in airway remodeling between subjects with severe and moderate asthma. *J Allergy Clin Immunol.* 2005;116(3):544-549.
79. Kaminska M, Foley S, Maghini K, Stoness-Bliss C, Coxson H, Ghezzoh H, et al. Airway remodeling in subject with severe asthma with or without chronic persistent airflow obstruction. *J Allergy Clin Immunol.* 2009;124(1):45-51. e1-4.
80. Ramos-Barbon D, Fraga-Iriso R, Brienza NC, Montero-Martinez G, Vera-Hernando H, Olivenstein R, et al. T cell localize with proliferating smooth muscle  $\alpha$ -actin+ cell

- compartment in asthma. *Am J Respir Crit Care Med.* 2010;182(3):317-24.
81. Dunnill MS, Massarella GR, Anderson JA. A comparison of quantitative anatomy of bronchi in normal subjects, in status asthmaticus, in chronic bronchitis and emphysema. *Thorax.* 1971;24(2):174-9.
82. Benayoun L, Druilhe A, Dombret MC, Aubier M, Pretolani M. Airway structural alterations selectively associated with severe asthma. *Am J Respir Crit Care Med.* 2003;167(10):1360-8.
83. Bai TR, Cooper J, Koelmeyer T, Pare D, Weir TD. The effect of age and duration of disease on airway structure in fatal asthma. *Am J Respir Crit Care Med.* 2000;162(2 Pt 1):663-9.
84. Ebina M, Yaegashi H, Chiba R, Takahashi T, Motomiya M, Tanemura M. Hyperreactive site in the airway tree of asthmatic patients revealed by thickening of bronchial muscles: a morphometric study. *Am Rev Respir Dis.* 1990;141(5 Pt 1):1327-32.
85. Saetta M, Di Stefano A, Rosina C, Thiene G, Fabbri LM. Quantitative structural analysis of peripheral airways and arteries in sudden fatal asthma. *Am Rev Respir Dis.* 1991;143(1):138-43.
86. Ebina M, Takahashi T, Chiba T, Motomiya M. Cellular hypertrophy and hyperplasia of airway smooth muscles underlying bronchial asthma: a 3-D morphometric study. *Am Rev Respir Dis.* 1993;148(3):720-6.
87. James AL, Wenzel S. Clinical relevance of airway remodelling in airways disease. *Eur Respir J.* 2007; 30(1):134-55.
88. Goldsmith AM, Bentley JK, Zhou L, Jia Y, Bitar KN, Fingar DC, et al. Transforming growth factor- $\beta$  induces airway smooth muscle hypertrophy. *Am J Respir Cell Mol Biol.* 2006;34(2):247-54.
89. Xie S, Sukkar MB, Issa R, Khorasani NM, Chung KF. Mechanisms of induction of airway smooth muscle hyperplasia by transforming growth factor-
- $\beta$ . *Am J Physiol Lung Cell Mol Physiol.* 2007; 293(1):L245-53.
90. Halwani R, Al-Muhsen S, Al-Jahdali H, Hamid Q. Role of transforming growth factor- $\beta$  in airway remodeling in asthma. *Am J Respir Crit Care Med.* 2011;44(2):127-33.
91. Salter B, Pray C, Radford K, Martin JG, Nair P. Regulation of human airway smooth muscle cell migration and relevance to asthma. *Respir Res.* 2017;18(1):156.
92. Parameswaran K, Cox G, Radford K, Janssen LJ, Sehmi R, OByrne PM. Cysteinyl leukotrienes promote human airway smooth muscle migration. *Am J Respir Crit Care Med.* 2002;166(5):738-42.
93. Goncharova EA, Billington CK, Irani C, Vorotnikov AV, Tkachuk AV, Penn RB, et al. Cyclic AMP-mobilizing agents and glucocorticoids modulate human smooth muscle cell migration. *Am J Respir Crit Care Ned.* 2003;29(1):19-27.
94. Parameswaran K, Radford K, Fanat A, Stephen J, Bonnans C, Levy BD, et al. Modulation of human airway smooth muscle migration by lipid mediators and Th2 cytokines. *Am J Respir Cell Mol Biol.* 2007;37(2):240-7.
95. Hecker L, Vittal R, Jones T, Jagirdar R, Luckhart TR, Horowitz JC, et al. NADPH oxidase-4 mediates myofibroblast activation and fibrogenic responses to lung injury. *Nat Med.* 2009;15(9):1077-81.
96. Sutcliffe A, Hollins F, Gomez E, Saunders R, Doe C, Cooke M, et al. Increased nicotinamide adenine dinucleotide phosphate oxidase 4 expression mediates intrinsic airway smooth muscle hypercontractility in asthma. *Am J Respir Crit Care Med.* 2012;185(3):267-74.
97. Sturrock A, Huecksteadt TP, Sanders K, Murphy TM, Chitano P, Wilson K, et al. Nox4 mediates TGF-1-induced retinoblastoma protein phosphorylation, proliferation, and hypertrophy in human airway smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol.* 2007;292(6):L1543-55.

98. Berair R, Saunders R, Brightling CE. Origin of airway smooth muscle mass in asthma. *BMC Med.* 2013;11:145.
99. Wicks J, Haitchi HM, Holgate ST, Davies DE, Powell RM. Enhanced upregulation of smooth muscle related transcripts by TGF $\beta$ 2 in asthmatic (myo)fibroblasts. *Thorax.* 2006;61(4):313-9.
100. Hackett T-L, Warner SM, Sefanowicz D, Shaheen F, Pechkovsky DV, Murray LA, et al. Induction of epithelial-mesenchymal transition in primary airway epithelial cells from patients with asthma by transforming growth factor- $\beta$ 1. *Am J Respir Crit Care Med.* 2009;180(2):122-33.
101. Takeda N, Sumi Y, Prefontaine D, Al Abri J, Al Heilay N, Al-Ramli W, et al. Epithelium-derived chemokines induce airway smooth muscle migration. *Clin EXP Allergy.* 2009;39(7):1018-26.
102. Chang Y, Al-Alwan L, Risso PA, Roussei L, Rousseau S, Halayko AJ, et al. Th17 cytokines induce human airway smooth muscle cell migration. *J Allergy Clin Immunol.* 2011;127(4):1046-1053.e1-2.
103. Redhu NS, Shan L, Movassagh H, Gounni AS. Thymic stromal lymphopoitin induces migration in human airway smooth muscle cells. *Sci Rep.* 2013;3:2301.
104. Govindaraju V, Michoud MC, Al-Chalabi M, Ferrano P, Powell WS, Martin JG. Interleukin-8: novel roles in human airway smooth muscle cell contraction and migration. *Am J Physiol.* 2006; 291(5):C957-65.
105. Joubert P, Lajoie-Kadoch S, Labonte I, Gounni AS, Maghni K, Wellemans V, et al. CCR3 expression and function in asthmatic airway smooth muscle cells. *J Immunol.* 2005;175(4):2702-8.
106. Kaur D, Saunders R, Berger P, Siddiqui S, Woodman L, Wardlaw A, et al. Airway smooth muscle and mast cell-derived CC chemokine ligand 19 mediate airway smooth muscle migration in asthma. *Am J Respir Crit Care Med.* 2006;174(11):1179-88.
107. Al-Alwan LA, Chang Y, Mogas A, Halayko AJ, Baglolle CJ, Martin JG, et al. Differential roles of CXCL2 and CXCL3 and their receptors in regulation normal and asthmatic airway smooth muscle cell migration. *J Immunol.* 2013;191(5):2731-41.
108. Ghaffer O, Hamid Q, Renzi PM, Allarkhverdi Z, Molet S, Hogg JC, et al. Constitutive and cytokine-stimulation expression of eotaxin by human airway smooth muscle cells. *Am J Respir Crit Care Med.* 199;156(6):1933-42.
109. Chung KF, Patel HJ, Fadlon EJ, Rousell J, Haddad EB, Jose PJ, et al. Induction of eotaxin expression and release from human airway smooth muscle by IL-1 and TNF: effects of IL-10 and corticosteroids. *Br J Pharmacol.* 1999;127(5):1145-50.
110. Hirst SJ, Hallsworth MP, Peng Q, Lee TH. Selective induction of release of eotaxin by interleukin-13 or IL-4 in human airway smooth muscle cells is synergistic with interleukin-1 and is mediated by the interleukin-4 receptor-chain. *Am J Respir Crit Care Med.* 2006;165(8):1161-71.
111. Palgan K, Bartuzi Z. Angiogenesis in bronchial asthma. In *J Immunopathol.* 2015;28(3):415-420.
112. Vrugt B, Wilson S, Bron A, Holgate ST, Djukanovic R, Aalbers R. Bronchial angiogenesis in severe glucocorticoid-dependent asthma. *Eur Respir J.* 2000;15(6):1014-21.
113. Wilson JW, Robertson CF. Angiogenesis in paediatric airway disease. *Paediatr Respir Rev.* 2002; 3(3):219-29.s
114. Tanaka H, Yamada G, Saikai T, Hashimoto M, Tanaka S, Suzuki K, et al. Increased airway vascularity in newly diagnosed asthma using a high-magnification bronchovideoscope. *Am J Respir Crit Care Med.* 2003;168(12):1495-9.
115. Barbato A, Turato G, Baraldo S, Bazzan E, Calabrese F, Panizzolo C, et al. Epithelial damage and angiogenesis in the airways of children with asthma. *Am J Respir Crit Care Med.* 2006;174(9):975-81.

116. Chetta A, Zanini A, Torre O, Olivieri D. Vascular remodelling and angiogenesis in asthma: morphological aspects and pharmacological modulation. *Inflamm Allergy Drug Targets.* 2007;6(1):41-5.
117. Hashimoto M, Tanaka H, Abe S. Quantitative analysis of bronchial wall vascularity in the medium and small airways of patients with asthma and COPD. *Chest.* 2005;127(3):965-72.
118. Nagy JA, Benjamin L, Zeng H, Dvorak AM, Dvorak HF. Vascular permeability, vascular hyperpermeability and angiogenesis. *Angiogenesis.* 2008;11(2):109-19.
119. Keglowich LF, Borge P. The three A's in asthma - airway smooth muscle, airway remodeling & angiogenesis. *Open Respir Med.* 2015;9:70-80.
120. Li X, Wilson JW. Increased vascularity of the bronchial mucosa in mild asthma. *Am J Respir Crit Care Med.* 1997;156(1):229-33.
121. Orsida BE, Li X, Hickey B, Thien F, Wilson JW, Walters EH. Vascularity in asthmatic airways: relation to inhaled steroid dose. *Thorax.* 1999;54(4):289-95.
122. Tuder RM, Yun JH. Vascular endothelial growth factor of the lung: friend or foe. *Curr Opin Pharmacol.* 2008;8(3):255-60.
123. Zanini A, Chetta A, Imperatori AS, Spanevello A, Olivieri D. The role of bronchial microvasculature in the airway remodelling in asthma and COPD. *Respir Res.* 2010;11(1):132.
124. Alapappan VK, de Boer WI, Mistra VK, Mooi WJ, Sharma HS. Angiogenesis and vascular remodeling in chronic airway disease. *Cell Biochem Biophys.* 2013;67(2):219-34.
125. Kanazawa H. VEGF, angiopoietin-1 and -2 in bronchial asthma: new molecular targets in airway angiogenesis and microvascular remodeling. *Rec Patents Inflam Allergy Drug Disc.* 2007;1(1)1-8.
126. Hoshino M, Takahashi M, Aoike N. Expression of vascular endothelial growth factor, basic fibroblast growth factors and angiogenin immunoreactivity in asthmatic airway. *J Allergy Clin Immunol.* 2001; 107(2):295-3001.
127. Yao X, Wang W, Li Y, Huang P, Zhang Q, Wang J, et al. IL-25 induces airway angiogenesis and expression of multiple angiogenic factors in murine model. *Respir Res.* 2015;16(1):39.
128. Wilson JW, Hii S. The importance of airway microvasculature in asthma. *Curr Opin Allergy Clin Immunol.* 2006;6(1):51-5.
129. Walters EH, Soltani A, Reid DW, Ward C. Vascular remodelling in asthma. *Curr Opin Allergy Clin Immunol.* 2008;8(1):39-43.
130. Shulman K, Rosen S, Tognazzi K, Manseau EJ, Brown LF. Expression of vascular permeability factor (VPF/VEGF) is altered in many glomerular diseases. *J Am Cos Nephrol.* 1996;7(5):661-6.
131. Meyer N, Christoph J, Makrinioti H, Indermitte P, Rhyner C, Soyka M, et al. Inhibition of angiogenesis by IL-32: Possible role in asthma. *J Allergy.* 2012;129(4):964-73.e7.
132. Bailey SR, Boustany S, Burgess JK, Hirst SJ, Sharma HS, Simcock DE, et al. Airway vascular reactivity and vascularization in humal chronic airway disease. *Pulm Pharmacol Ther.* 2009;22(5):417-25.
133. Simcock DE, Kanabar V, Clarke GW, Mahn K, Karner C, O'Connor BJ, et al. Induction of angiogenesis by airway smooth muscle from patients with asthma. *Am J Respir Crit Care Med.* 2008; 178(5):460-8.
134. Van Eerdewegh P, Little RD, Dupuis J, Del Mastro RG, Falls K, Simon J, et al. Association of ADAM33 gene with asthma and bronchial hyperresponsiveness. *Nature.* 2002;418(6896):426-30.
135. Rao SP, Sikora L, Hosseinkhani MR, Pinkerton KE, Sriramara P. Exposure to environmental tobacco smoke induces angiogenesis and leukocyte trafficking in lung microvessel. *Exp Lung Res.* 2009; 35(2):119-135.

136. McKnight CG, Potter C, Finkelman FD. IL-4R $\alpha$  expression by airway epithelium and smooth muscle accounts for nearly all airway hyperresponsiveness in murine allergic airway disease. *Mucosal Immunol.* 2019;13(2):283-92.
137. Gaevreu GM, Sehmi R, Ambrose CS, Griffiths JM. Thymic stromal lymphopoietin: its role potential as a therapeutic agent in asthma. *Expert Opin Ther Targets.* 2020;24(8):777-92.
138. Ingram JL, Kraft M. IL-13 in asthma and allergic disease: asthma phenotypes and targeted therapies. *J Allergy Clin Immunol.* 2012;130(4):829-42.
139. Brooks GD. Updated evaluation of dupilumab in the treatment of asthma: patient selection and reported outcomes. *Ther Clin Risk Manag.* 2020;16:181-7.
140. Busse WW, Maspero JF, Lu Y, Ruddy M, Graham NMH, Teper A. Efficacy of dupilumab on clinical outcomes in patients with asthma and perennial allergic rhinitis. *J Allergy Clin Immunol.* 2020;125(5):565-76.e1.
141. Regeneron Pharmaceutical, Inc.
142. Matera MG, Rogliani P, Calzetta L, Cazzola M. TSLP inhibitors for asthma: current status and future prospects. *Drugs.* 2020;80(5):449-459.
143. Corren J, Parnes J, Wang L, Mo M, Roseti SL, Griffiths JM, et al. Tezepelumab in adults with uncontrolled asthma. *N Engl J Med.* 2017;377(10):936-46.
144. Pham TH, Ren P, Parnes JR, Griffiths JM. Tezepelumab reduces multiple key inflammatory biomarkers in patients with severe uncontrolled asthma in the Phase 2b PATHWAY study. *Am J Respir Crit Care Med.* 2019;119:A2679.
145. Corren J, Garcia Gil E, Parnes J, Pham T, Griffiths JM. Tezepelumab treatment effect on annualized rate of exacerbation by baseline biomarkers in uncontrolled severe asthma: phase 2b PATHWAY study. *Am J Respir Crit Care Med.* 2019;100:A2621.
146. Menzie-Gow A, Colice G, Griffiths JM, Almqvist G, Ponnarambil S, Kaur P, et al. NAVIGATOR: a phase 3 multicentre, randomized, double-blind, placebo-controlled, parallel group trial to evaluate the efficacy and safety of tezepelumab in adults and adolescents with severe, uncontrolled asthma. *Respir Res.* 2020;21(1):266.
147. Syabbalo N. The role of alarmin cytokines in the pathogenesis of severe uncontrolled asthma. *Ann Clin Med Res.* 2021;2(1):1022.
148. Zuyderduyn S, Sukkar MB, Fust A, Dhaliwal S, Burgess JK. Treating asthma means treating airway smooth cells. *Eur Respir J.* 2008;32(2):265-74.
149. Tan LD, Yoneda KY, Louie S, Hogarth DK, Castro M. Bronchial thermoplasty: A decade experience: state of the art. *J Allergy Clin Immunol in Pract.* 2019;7(1):71-80.
150. Pretolani M, Dombret MC, Thabut G, Knap D, Hamidi F, Debray MP, et al. Reduction of airway smooth muscle mass by bronchial thermoplasty in patients with severe asthma. *Am J Respir Crit Care Med.* 2014;190(12):1452-4.
151. Chupp G, Laviolette M, Cohn L, McEvory C, Bansal S, Shifren A, et al. Long-term outcomes of bronchial thermoplasty in subjects with severe asthma: a comparison of 3-year follow-up results from two prospective multicenter studies. *Eur Respir J.* 2017;50(2):1700017.
152. d'Hooghe JNS, Ten Hacken NHT, Weersink EJM, Sterk PJ, Annema JT, Bonta PI. Emerging understanding of the mechanism of action of bronchial thermoplasty in asthma. *Pharmacol Ther.* 2018;181:101-7.
153. Thomson NC. Recent developments in bronchial thermoplasty for severe asthma. *J Asthma Allergy.* 2019; 12:375-387.
154. Syabbalo N. Bronchial thermoplasty in the treatment of severe asthma. *J Pulmonol Res Rep.* 2020;2(2):3-9.
155. Castro M, Rubin AS, Laviolette M, Fiterman J, De Andrade Lima M, Shah PL, et al. Effectiveness and

- safety of bronchial thermoplasty in the treatment of severe asthma: a multicenter, randomized, double-blind. Sham-controlled clinical trial. Am J Respir Crit Care Med. 2010;181(2):116-24.
156. U.S. Food and Drug Administration, Alair Bronchial Thermoplasty system: Alair catheter and Alair RF controller; 2010.

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